

TOXIC IMPACT OF PESTICIDES ON THE MORPHOLOGICAL CHARACTERISTICS OF BLOOD CELLS OF FISH *Channa punctatus* (BLOCH)

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ABSTRACT

Pesticide is an agrochemical generally used for controlling the pests of agricultural products but when it get mixed with water resources through agricultural lands, it affects the river water and the organisms as well as the whole river water ecosystem. This affect being an adverse one can change the physiological, behavioral and biochemical properties of aquatic organisms. In the present study an attempt has been made to find out the toxic effect of agrochemical "trichlorofan" on the morphological structure of Blood cells of fish *Channa punctatus* (Bloch). The morphological characteristics were investigated after exposing the fish to a pesticide Trichlorofan, which is generally used for pest control by the farmers. Blood samples were collected from both control i.e. untreated and treated fish after 24,48,72,96,120,148 and 172 hours of exposure and blood films were prepared for morphological study. Significant changes in the morphology of blood cells were obtained between treated groups of fish when compared with the control one.

KEYWORDS: Agrochemical, Blood, Biochemical, Morphology, Pesticide

Environmental pollution is one of the most serious problems facing humanity and other life forms on our planet today. Some of these pollutants are responsible for various acute and chronic diseases, such as skin burns and rashes, bone abnormalities, lung and digestive system disorders in aquatic and surrounding land fauna. (John, 1990).

Pesticide is an agrochemical generally used for controlling the pests of agricultural products but when it get mixed with water resources through agricultural lands, it affects the river water and the organisms as well as the whole river water ecosystem. This affect being an adverse one can change the physiological, behavioral and biochemical properties of aquatic organisms. Fishes are aquatic and poikilothermic animals. Hence, their existence and performance is dominated by the quality of their environment. All species of fish perform best under certain optimal conditions, but the amplitude of such conditions is quite narrow. A fish's survival in the face of environmental stress thus, depends up on its ability to adjust its physiological processes so as to maintain relatively constant internal body chemistry. Any stress requiring an adjustment in excess of ability to accommodate will be lethal or will result in disease (Wedemeyer,1970, Wedemeyer and Wood,1974).

Haematological indices are important parameters to evaluate the general physiological

status of fishes and may be considered as stress indicators for estimation of the response reactions of the fish to various environmental conditions (Docan et al.,2010). Besides, haematological tests can also provide important information on the erythropoietic condition (Rehulka and Adamec, 2004). it may be considered useful in assessing the health of fish subjected to changing environmental conditions (Blaxhall,1972 and Nair et al, 1984) and have proven useful in monitoring stress responses as bio-indicators (Bridges et al,1976; Soivio & Oikari,1976;Warner & Wiliams,1977; Agarwal and Shrivastava 1980;Folmar,1993;Gill & Eppele,1993; Caruso et al ,2005;Remya et al,2008 and Ramesh and Saravanan,2008,Singh and Tandon,2009).

Several studies have been conducted regarding the reactions and modifications of the haematological indices in response to various stress factors (Gbore et al., 2006). Montero et al. (1999) reported increases in hematocrit value, haemoglobin amount and red blood cell count, upon short term density stress, suggesting a strategy to increase the ability of oxygen transportation in the blood during periods of metabolic breakdown. Significant increases in hematocrit, haemoglobin, and MCHC during high stocking density environment have been reported in *Salmo salar* (Kjartansson et al. 1988), while in *Carassus auratus* both the hematocrit and the amount of haemoglobin depleted with the

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increasing stocking density (Burton and Murray,1979).

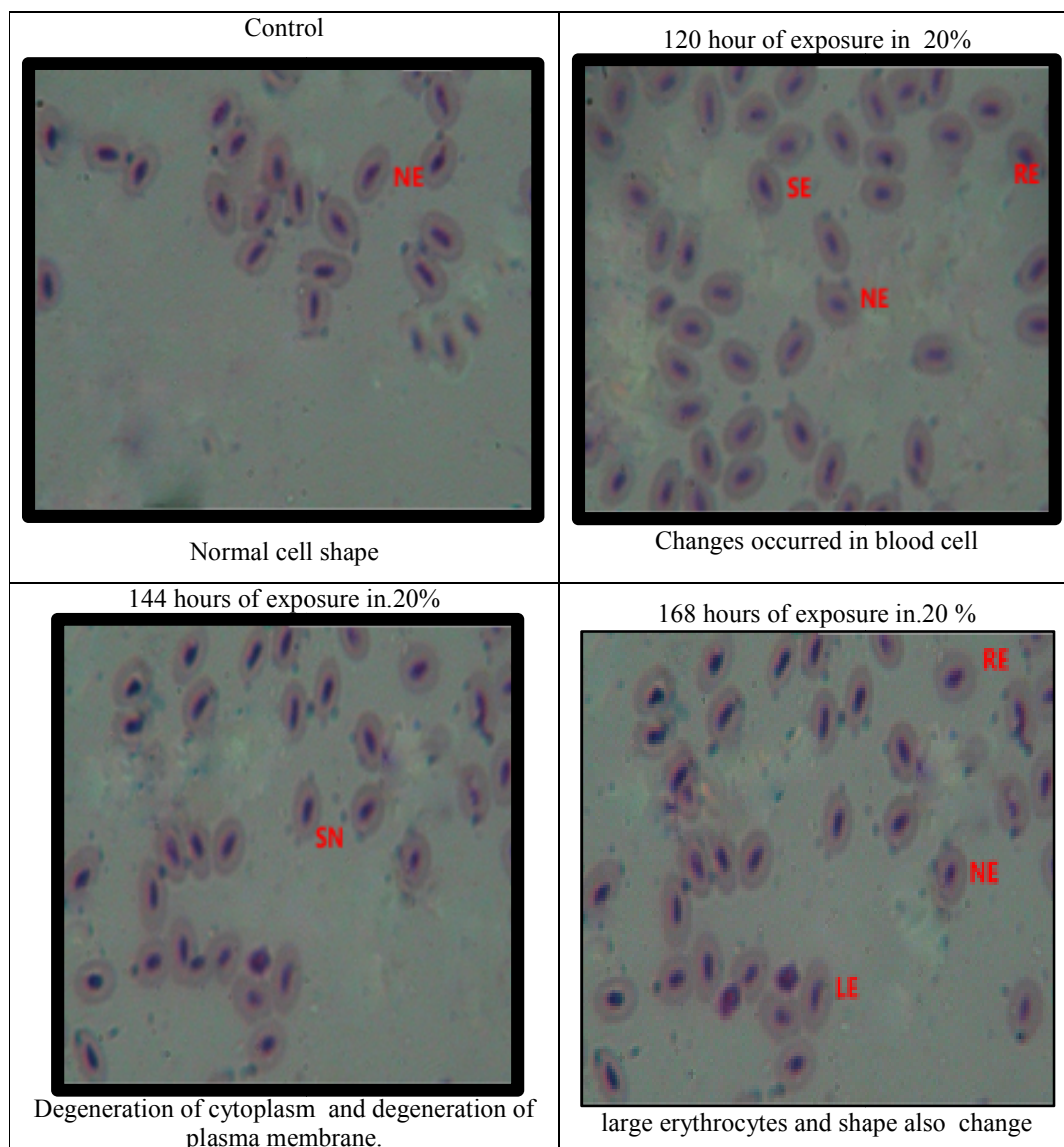
MATERIALS AND METHODS

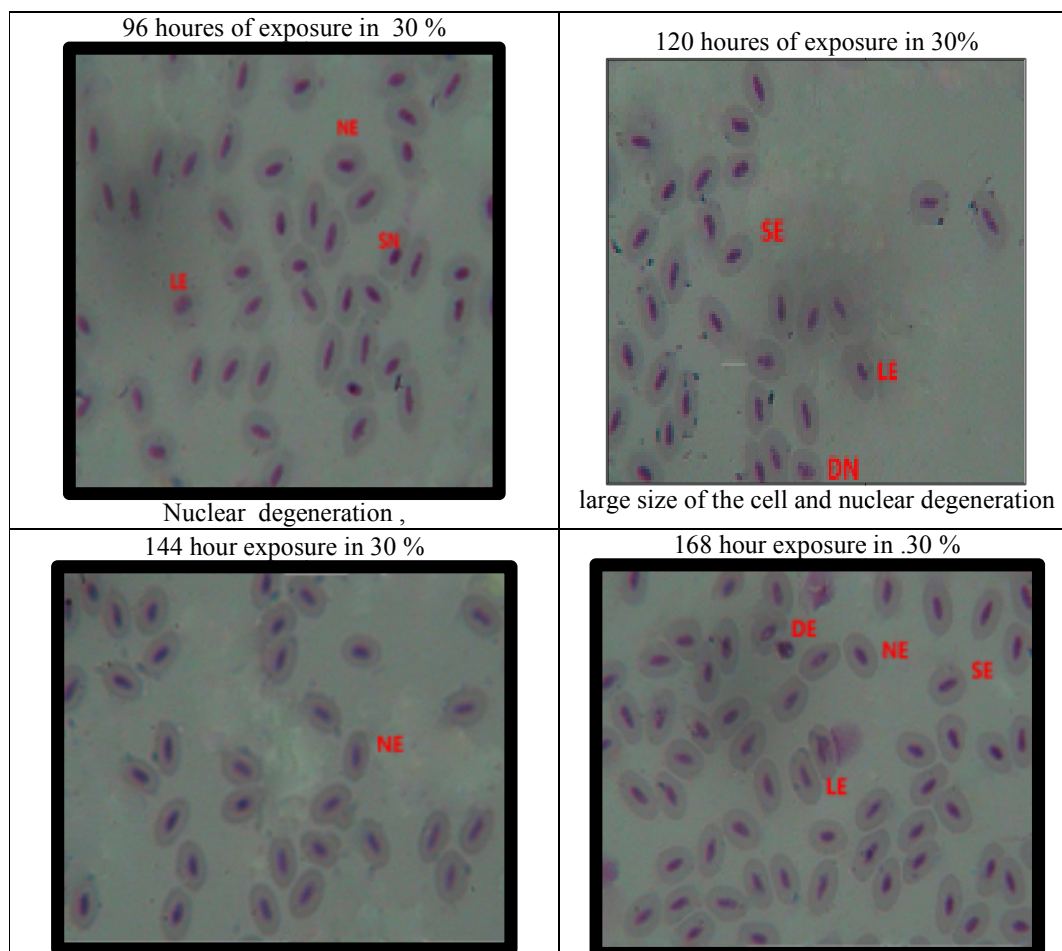
Collection and Acclimatization of Fishes

Live, healthy *Channa punctatus* in the same size range were collected from local streams and acclimatized under normal laboratory conditions for 15 days. For the exposure of fishes, different concentration ranges (10%,20%, and 30%) were taken after dilution with normal tap water and Normal tap water served as control. The fishes were

divided into 5 groups consisting of 20 fishes each. All were individually exposed to Tap water (Control) and low concentrations (10%,20%,and 30%) of agrochemical in 20 liters glass aquaria. Feeding of fishes and aeration of the tanks were done uniformly throughout the experiment. Blood samples were collected separately from 4 live fishes from each group at 24,48,72,96,120,144 &168 hours by severing the caudal peduncle. A thin blood film is prepared by spreading a small drop of blood uniformly. After staining with Leshmania stain the blood film is used to determine the cell type characteristics and number.

Figure 1to 8 shows the morphological changes in blood cell of fish *Channa punctatus* (Bloch)





RESULTS

After exposure in different concentration of agrochemical many morphological changes was observed .during this it No any changes were observed in 10% concentration of after the exposure while during the Exposure to 20 % effluent revealed the presence of anisocytosis at 96 hours [plate 1], damaged erythrocytes at 120 hours [plate 2], anisocytosis , deformed erythrocytes and large bulged nucleus in some cells at 144 hours [V-3(E)],clumped erythrocytes at 176 hours [plate 3], deformed erythrocytes at 120 hours [plate 4] followed by deformed cells clumped erythrocytes and cell lysis at 144 and 168 hours [plate 5].Exposure to 30 % effluent revealed, anisocytosis and karyorhexis (nuclear degeneration) in the first 72 hours plate 6&2 (G).At 72 hours, the cells looked pale with a large nucleus marking the onset of cellular degeneration [plate 7].This was followed by karyorhexis and

cytoplasmic degeneration at 96 hours [plate 8]and large, clumped erythrocytes and cell lysis at 120 to 168 hours [plate 9].

Presence of anisocytosis and poikilocytosis in the present case clearly indicates toxic effects agrochemical on the red cell morphology. Structural defects and changes in surface shapes of erythrocytes have been reported by Koc et al. (2008) from Endosulfan and Malathion exposed rats. Changes in the erythrocyte profile were also noticed in fishes by Benarji and Rajendranath(1990) in presence of Dichlorvos, Tavares et al. (1999) to Trichlorphon, Khattak and Hafeez (1996) to Malathion, Singh and Srivastava (1994) to Formothion and Sampath et al. (1993) Ekolux organophosphorus preparation. Deformations and increase in the cellular size of the red blood cells were also observed in the present case prior to clumping. Similar changes have been observed by Tripathi and Shrivastava (2010) in rats

post exposure to Chlorpyrifos. Comelekoglu et al. (2000) stated that some pesticides may provoke the alterations in size and surface shapes of erythrocytes. Nikimma (1992) suggested that toxic materials directly or indirectly damage the membrane structure, ion permeability and cell metabolism of erythrocytes thus may cause morphologically damaged erythrocyte formation. Vives et al. (1999) explained that the expansion of membrane increases the area/volume proportion and could allow the swelling of the cell, thus, reaching the largest volume before the lysis. Swelling of red blood cells as reflected by the increased mean corpuscular volume (MCV) has been attributed to the increase in the activity (Soivio et al. 1974). An increase of erythrocyte size (MCV) has been associated with several factors but it is generally considered as a response to stress (Weber 1982).

REFERENCES

- Agrawal S.J. and Srivastava A.K., 1980. Effect of Endosulfan on certain aspects of haematology of the fish, *Channa punctatus*. *Toxicology*, **17**: 97.
- Banerji G. and Rajendranath T., 1990. Haematological changes induced by an organophosphorus insecticide in a freshwater fish *Clarias batrachus* (Linnaeus). *Trop. Freshwater. Biol.*, **2**: 197-202
- Blaxhall P.C., 1972. The hematological assessment of the health of freshwater fish. *J. Fish Biol.*, **4**: 593-6057 Nair et al, 1984
- Bridges D.W., Cech J.J. and Pedro D.N., 1976. Seasonal hematological changes in winter flounder, *Pseudopleuronectes americanus*. *Trans.Am.Fish Soc.*, **105**: 596-600.
- Burton C.B. and Murray S.A., 1979. Effect of density on gold fish blood. 1. *Haematol.*, **62A**: 555- 558.
- Caruso G., Genovese L., Maricchiolo G. and Modica A., 2005. Haematological, biochemical and immunological parameters as stress indicators in *Dicentrarchus labrax* and *Sparus aurata* farmed in off-shore cages. *Aquaculture International* , **13**:67–73.
- Docan A., Cristea V., Grecu I. and Dediu L., 2010. Haematological response of the European catfish, *Silurus glanis* reared at different densities in “flow-through” production system. *Archiva Zootechnica*, **13**(2):63-70.
- Folmar L.C., 1993. Effects of chemical contaminants on blood chemistry of teleost fish: a bibliography and synopsis of selected effects. *Environ. Toxicol. Chem.*, **12**:337-375.
- Gbore F.A., Oginni O., Adewole A.M. and Aladetan J.O., 2006. The effect of transportation and handling stress on hematology and plasma biochemistry in fingerlings of *Clarias gariepinus* and *Tilapia zillii*. *World J. Agric. Sci.*, **2**(2): 208-212 (Cited by Docan et al, 2010).
- Gill T.S. and Epple A., 1993. Stress-related changes in hematological profile of the American eel (*Anguilla rostrata*). *Ecotoxicol. Environ. Safety*, **25**: 227–235.
- John De Z., 1990. Handbook of drinking water quality standards and control. *Van Nostra and Reinhold Publications*. New York. 34-151.
- Khattak I.U.D. and Hafeez M.A., 1996. Effect of malathion on blood parameters of the fish, *Cyprinion watsoni*. *Pak J. Zool.*, **28**: 45-49.
- Kjartansson H., Fivelstad S., Thomassen J.M. and Smith M.J., 1988. Effects of stocking densities on physiological parameters and growth of adult Atlantic salmon (*Salmo salar* L.) reared in circular tanks. *Aquaculture*, **73**: 261-274.
- Nikimma M., 1992. How does environmental pollution affect red blood cell function in fish ? *Aquat.Toxicol.*, **22**: 227-238.
- Ramesh M. and Saravanan M., 2008. Hematological and biochemical responses in freshwater fish *Cyprinus carpio* exposed to Chlorpyrifos. *Inter. Jour. Integrative Biol.*, **3**(1): 8017.
- Remyła S.R., Ramesh M., Sajwan K.S. and Kumar K.S., 2008. Influence of zinc on cadmium induced haematological and biochemical

- responses in a freshwater teleost fish *Catla catla*. *Fish Physiol. Biochem.*, 34: 169–174. DOI 10.1007/s10695-007-9157-2
- Sampath K., Velammal S. and Kennedy J.R., 1993. Haematological changes and their recovery in *Oreochromis mossambicus* as a function of exposure period and sublethal levels of Ekalux. *Acta hydrobiol.*, 35: 73-83.
- Singh N.N. and Srivastava A.K., 1994. Formation induced haematological changes in the freshwater Indian catfish *Heteropneustes fossilis*. *J.Ecotox.Environ.Monit.* 4:137-140.
- Singh B.P. and Tandon P.K., 2009. Effect of river water pollution on hematological parameters of fish, Wallago attu. *Res. Environ. Life Sci.*, 2(4):211-214. ISSN: 0974-4908
- Soivio A. and Oikari A., 1976. Hematological effects of stress on a teleost *Esox luciosus* L. *J.Fish Biol.*, 8:397-411.
- Tavares D.M., Martins M.L. and Nascimento K.S., 1999. Evaluation of haematological parameters in *Praractus mesopotamicos* Holmberg (Osteichthyes: Characidae) with *Argulus* sp. (Crustacea: Branchiura) infestation and treatment with Organophosphate. *Rev. Bras.*, 16:553-555.
- Tripathi S. and Shrivastava A.K., 2010. Nephrotoxicity induced by long-term oral administration of different doses of Chlorpyrifos. *Toxicol. Ind.Health.*, 26(7):439-447. DOI:10.1177/0748233710371110
- Vives M.A., Infante M.R., Garcia E., Selve C., Mangras M. and Vinardell M.P., 1999. Hemolytic action of anionic surfactants of the diacyl lysine type. *Chem. Biol. Interact.*, 118: 1-18
- Warner M.C. and Williams R.W., 1977. Comparisons between serum values of pond and intensive raceway cultured channel Catfish *Ictalurus punctatus* (Rafinesque). *J.Fish Biol.*, 11: 385-391.
- Weber R.E., 1982. Interspecific adaptation of haemoglobin function in fish to oxygen availability. In: Addink A.D. and Spronk N.(Eds.), *Exogenous and Endogenous Influences on Metabolic and Neural control*. Pergamon, Oxford, 87-101.
- Wedemeyer G.A. and Wood J.W., 1974. Stress as a predisposing factor in fish diseases. Federal Government Series: Fish Disease Leaflet – 38, U.S. Fish and Wildlife Service, USGS.
- Wedemeyer G., 1970. The role of stress in the disease resistance of fishes. P: 30-35 In: Snieszko, S.F. ed. *A symposium on diseases of fishes and shell fishes*. Spec. publ. 5, Am.Fish Soc., Washington, DC.